Bacteria Associated with the Spoilage of Salad, their resistotyping and Potential Public Health Implications

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Abstract—A study of bacteria associated with spoilage of various types of salads purchased from 3 different restaurants in Uyo metropolis was carried out between the months August and October, 2008 using standard microbiological procedures. The mean microbial load in vegetable salad was (3.5±0.25) x 10^7 cfu/g with the sample from food affairs having the highest load (3.7 x 10^7 cfu/g). The mean microbial load in garlic salad was (3.2±0.25) x 10^7 cfu/g also with the sample from food affairs having the highest load (3.6 x 10^7 cfu/g). However, fruit salad samples from Mr. Biggs had the highest microbial load (3.9 x 10^7 cfu/g) out of the mean value of (3.6±0.25) x 10^7 cfu/g. Analysis of the frequency of occurrence of the isolates revealed that Staphylococcus aureus (33%), Micrococcus spp. (20%), Bacillus spp. (20%), Proteus spp. (13%), Pseudomonas spp. (13%) and Escherichia coli (7%) were the most encountered organisms in both spoiling and fresh (control) salad. The resistotyping of 20 % of tested bacteria were resistant to 5 antibiotics. The presence of Escherichia coli and some potential pathogenic microorganisms in all the samples collected are indications of faecal contamination and unhygienic method of food handling. This suggests that salad should be properly managed during and after preparations to avoid outbreak of food borne diseases. Also the food handlers should be trained and educated on the importance of sanitary condition during food production.

Keywords—Bacteria, salad, spoilage, Staphylococcus species, Escherichia coli, resistotyping.

I. MATERIALS AND METHODS

A. Sources and Collection of Salad

Samples used in this work were purchased from 3 different fast food restaurants in Uyo metropolis. They were aseptically packaged prevent contaminations during transportation to laboratory. They were analyzed immediately to determine the bacterial load and diversities associated with fresh salad. The left over were repackaged and allowed to stand without refrigeration until the signs of spoilage were observed. Samples from the spoilt salad were re-analyzed as done to the fresh fruit (control). The rationale for determining the bacterial load and diversities in the fresh salad is to source track the bacterial isolates that would be determined later while spoiling.

B. Media, Reagents and Antibiotics

Media used which include Nutrient Agar (Oxoid, Cambridge, UK) macConkey Agar (Oxoid, Cambridge, UK), Eosin Methylene Blue Agar (Oxoid, Cambridge, UK) and reagents were purchased from Globask Scientific, Idumota, Lagos. The single antibiotic disks were purchased from Oxoid representative in Ogba, Ikeja, Lagos, Nigeria.

C. Determinations of Bacterial Load and Bacterial Diversities

Established 10 fold serial dilution methods of Collins and Lyne (1979) was employed to determine the total heterotrophic bacterial count and coliform count using nutrient agar and Eosin Methylene Blue Agar (Oxoid, Cambridge, UK) respectively. Determination of bacterial diversities in both fresh (control) and the spoiling salad was carried out using conventional methods described by Cheesbrough (2006) and Kit systems.

D. Resistotyping of the identified Bacterial Species:

Disc diffusion techniques as described by Cheesbrough (2006) was used for this test. The selected antibiotics were nalidixic acid, kanamycin, amoxicillin-clavulanic acid, oxytetracycline, cefotaxime and enrofloxacin.
II. RESULTS

Evidence of spoilage observed in the unrefrigerated salad several hours after purchase include change in colour, taste and obvious sign of primary decay in the sliced fruits.

A. Microbial Counts of the Salad Samples

The mean microbial load in vegetable salad was \((3.5\pm0.25) \times 10^7\) cfu/g with the sample from Fast Food FA having the highest load \((3.7 \times 10^7\) cfu/g). The mean microbial load in garlic salad was \((3.2\pm0.25) \times 10^7\) cfu/g also with the sample from Fast Food FA having the highest load \((3.6 \times 10^7\) cfu/g). However, fruit salad samples from Fast Food MB had the highest microbial load \((3.9 \times 10^7\) cfu/g) out of the mean value of \((3.6\pm0.25) \times 10^7\) cfu/g. The bacterial load in the freshly prepared salad was lower than the load observed in the spoiling salad. The least load was observed in garlic salad (Table 1 and 2).

B. Bacterial Diversities common in all the Salad during Spoilage and their Frequency of Occurrence

The bacterial *Staphylococcus aureus*, *Proteus vulgaris*, *Bacillus* spp., *Psedomonas* spp., *Micrococcus* spp., *Escherichia coli*. All the bacterial diversities were detected in both the freshly prepared and spoiling salad. The analysis of the frequency of occurrence of the isolates revealed that *Staphylococcus aureus* has the highest frequency of occurrence while the *Micrococcus* spp., *Bacillus* spp., *Proteus* spp., *Pseudomonas* spp. and *Escherichia coli* were the most encountered organisms (Table 3).

TABLE 1

<table>
<thead>
<tr>
<th>Sample type</th>
<th>Veg. (cfug(^{-1}) x 10(^3))</th>
<th>Control Veg. Salad (cfug(^{-1}) x 10(^3))</th>
<th>Sample type 2</th>
<th>Garlic Salad (cfug(^{-1}) x 10(^3))</th>
<th>Sample type 3</th>
<th>Fruit Salad (cfug(^{-1}) x 10(^7))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fast Food Outfits</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MB</td>
<td>3.5</td>
<td>1.7</td>
<td>3.3</td>
<td>1.3</td>
<td>3.9</td>
<td>1.4</td>
</tr>
<tr>
<td>FA</td>
<td>3.7</td>
<td>1.1</td>
<td>3.6</td>
<td>1.2</td>
<td>3.6</td>
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</tr>
<tr>
<td>OT</td>
<td>3.4</td>
<td>1.4</td>
<td>2.8</td>
<td>0.8</td>
<td>3.4</td>
<td>1.3</td>
</tr>
<tr>
<td>MEAN</td>
<td>3.5</td>
<td>1.4</td>
<td>3.2</td>
<td>1.1</td>
<td>3.6</td>
<td>1.4</td>
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</table>
### TABLE 2
**Coliform Counts from the Unrefrigerated Salad Undergoing Spoilage and Control (Freshly Prepared)**

<table>
<thead>
<tr>
<th>Fast Food Outlet</th>
<th>Sample 1</th>
<th>Sample 1 Control</th>
<th>Sample 2</th>
<th>Sample 2 Control</th>
<th>Sample 3</th>
<th>Sample 3 Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vegetable Salad</td>
<td>Veg Salad (cfug⁻¹) x 10⁵</td>
<td>Vegetable Salad (cfug⁻¹) x 10⁵</td>
<td>Fruit Salad (cfug⁻¹) x 10⁵</td>
<td>Fruit Salad (cfug⁻¹) x 10⁵</td>
<td>Garlic Salad (cfug⁻¹)</td>
<td>Garlic Salad (cfug⁻¹)</td>
</tr>
<tr>
<td></td>
<td>MB</td>
<td>FA</td>
<td>OT</td>
<td>x</td>
<td>MB</td>
<td>FA</td>
</tr>
<tr>
<td></td>
<td>1.8</td>
<td>2.7</td>
<td>2.1</td>
<td>2.2</td>
<td>1.8</td>
<td>2.7</td>
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<tr>
<td></td>
<td>0.7</td>
<td>1.1</td>
<td>1.2</td>
<td>1.0</td>
<td>0.7</td>
<td>1.1</td>
</tr>
<tr>
<td></td>
<td>2.0</td>
<td>2.3</td>
<td>1.5</td>
<td>1.9</td>
<td>2.0</td>
<td>2.3</td>
</tr>
<tr>
<td></td>
<td>1.0</td>
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<td>1.5</td>
<td></td>
<td>1.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>2.3</td>
<td></td>
<td>1.9</td>
<td></td>
<td>2.3</td>
<td></td>
</tr>
<tr>
<td></td>
<td>6</td>
<td></td>
<td>2.2</td>
<td></td>
<td>6</td>
<td></td>
</tr>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

### TABLE 3
**Frequency of Occurrence of Bacteria and Coliform in the Three Salad Samples.**

<table>
<thead>
<tr>
<th>S/N</th>
<th>Type of Organisms</th>
<th>Frequency of Occurrence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td><em>Staphylococcus</em> spp</td>
<td>33</td>
</tr>
<tr>
<td>2</td>
<td><em>Proteus</em> spp</td>
<td>20</td>
</tr>
<tr>
<td>3</td>
<td><em>Bacillus</em> spp</td>
<td>20</td>
</tr>
<tr>
<td>4</td>
<td><em>Pseudomonas</em> spp</td>
<td>13</td>
</tr>
<tr>
<td>5</td>
<td><em>Escherichia coli</em></td>
<td>7</td>
</tr>
<tr>
<td>6</td>
<td><em>Micrococcus</em> spp</td>
<td>7</td>
</tr>
<tr>
<td></td>
<td></td>
<td>100</td>
</tr>
</tbody>
</table>

### TABLE 4
**Resistotyping of the Bacterial Isolates**

<table>
<thead>
<tr>
<th>NA</th>
<th>C</th>
<th>AUG</th>
<th>OT</th>
<th>CTX</th>
<th>ENF</th>
</tr>
</thead>
<tbody>
<tr>
<td>3S 3I 5R</td>
<td>8S 1I 2R</td>
<td>5S 6R</td>
<td>8S 3R</td>
<td>2S 1R 8R</td>
<td>100S</td>
</tr>
<tr>
<td>27.7S</td>
<td>72.8S</td>
<td>45.5S</td>
<td>72.8S</td>
<td>18.2S</td>
<td></td>
</tr>
<tr>
<td>27.7I</td>
<td>9I</td>
<td>55.5R</td>
<td>27.2R</td>
<td>9I</td>
<td></td>
</tr>
<tr>
<td>45.6R</td>
<td>18.2R</td>
<td>72.8R</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

20% were Resistant to 5 Antibiotics

Key: NA= nalidixic acid, C= kanamycin, AUG= amoxicillin-clavulanic acid, OT=oxytetracycline, CTX= cefotaxime, ENF= enrofloxacin
III. DISCUSSION

The salad analysed in this study is an important food supplement because it is made from various vegetables and fruits which includes cabbage, white onion, carrot, cucumber, green peas, lettuce, spinach and cream salad. The consumption of spoil or contaminated salad will pose a greater danger to health because of the uncooked nature by which the salad is usually served. Since the salad samples analysed in this study indicated the presence of some potential pathogenic microorganisms, the knowledge of the bacterial flora of salad is necessary in order to effectively safeguard against the consumption of contaminated salad.

Analysis of the frequency of occurrence of the isolates in this study revealed that *Staphylococcus aureus*, *Micrococcus spp*, *Bacillus spp*, *Proteus spp*, *Pseudomonas spp*, *Escherichia coli* were the most encountered organisms. Deng *et al.*, (1998) emphasized the ability of this bacterium to survive in dry foods, with a wide range of aw and pH values, particularly at refrigeration temperatures. Abdul Raouf *et al.*, (1993) have reported that populations of *E. coli* O157: H7 (10⁵ cfu/g) inoculated in ground, roasted beef salads containing up to 40 % mayonnaise and held at 5 °C showed no changes even after 72 hr of incubation. *Staphylococcus aureus* is distributed in the environment and their primary inhabitant is the anterior nares and skin of man and animals (Adegoke and Komolafe, 2009). *Bacillus spp* can grow in food and produce enterotoxins that cause food poisoning. Both bacterial species are widely distributed in the soil, they often contaminated vegetable and fruits. Most of these organisms have been implicated in septicaemia (Komolafe and Adegoke, 2008).

Other organisms found include proteus sp and micrococcus sp. The presence of *Escherichia coli* in food, water and fruits is an indication of faecal contamination resulting from unhygienic method of food handling. The occurrence of pseudomonas sp in salad also can constitute a public health hazard, especially because of their high resistance to antibiotics. This types of multirdrug resistance of 20 % to 5 last line antibiotics make the isolated bacteria of high epidemiological importance (Adegoke and Komolafe, 2009).

IV. CONCLUSION AND RECOMMENDATION

This study has shown that salad although very delicious and nutritive do harbour microorganisms including pathogenic microorganism. The activities of microorganisms have been reported as the main cause of salad spoilage. The source of the microorganisms isolated in this study is suspected to come from the poor handled conditions and their proliferation enhanced.

Based on the result of this study, there is need that ready to eat food should be serve immediately preparation because result of this analysis has shown that some microorganisms can grow and multiply in the food item even under refrigeration. Producers of salad should make sure that the condiments used for the production of salad are of recommended microbiological standards. It is also important that food handlers are trained and educated on the importance of sanitary condition during food production.

REFERENCES


